

In Vivo Quantification of Integrin-Targeted and Protease-Activated Imaging Agents in Response to Anti-Angiogenic Therapy using Quantitative Fluorescence Tomography

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1 Abstract

Integrins are transmembrane cell surface receptors which mediate signal transduction, cell-to-cell interaction and cell-to-extracellular mark adhesion, key processes involved in angiogenesis, tumorigenesis and metastasis. Integrins have thus been halled as clinically-relevant biomarkers of pathological conditions such as inflammation and tumor progression. The integrin a_0 , is significantly upergulated in tumor cells and activated endothelial cells during neangingenesis. To date, it has not been possible to strictly quantify integrin levels in vivo with existing optical imaging technologies. What is more, the simultaneous quantification and in vivo localization of ritegrins and of a distinct biomarker has proved unfeasible. The aim of this study was to simultaneously and non-invasively image and quantify signal of α_0 , β_1 seceptor binding and the signal of a cathepian-cativable imaging agent using a specific, targeted near-infraerd (NIRs) flourescence agent and Fluorescence Molecular Tomography – or FMT^{IM} (FMT 2500^{IM} Quantitative Tomography System, PerkinElimer). We developed a fluorescence imaging agent (IntegriSensei)», PerkinElimer) in vivo detection of α , β_2 , suggest of molecular weight peptidominetic antagonist coupled to a NIR fluorochrome. The dissociation constant Kd, as determined by binding to α , β_3 -overgressing HEXC93 cells, was found to be 4.5 ± 10 nM. The pharmacokinetic profile was assessed in mice by measuring plasma fluorescence at different times after intravenous injection with the agent and found to fit a two-compartmental model and calculated to be $(1)_{1/2} = 0$ min and $(2)_{1/2} = 0$ min min. Integrine expression in tumors was quantified in both mouse breast tumors and human mabdomyosarcoma tumor xenografts implanted in nude mice, and the quantified fluorescent signal strongly correlated with tumor size (7–0.57). In a fluoridity, this agent can develop the control of the size of distribution reflecting underlying differences in integring and cathegation loopy during

2 Integrin-targeted Imaging Agent

The integrin-targeted agent (IntegriSense® 680, PerkinEtner) was synthesized by converting the small molecule, non-peptide a.b, a rhazgonist, compound of (Coleman et al.), to the 3-vano derivative, reducing the derivative to the 3-aminomethyl analog and reacting the resulting compound with VivOTag¹⁰ 680 (PerkinEtner), an amine-reactive near-infra-red fluorochrome designed to allow maximal bassor penetration and minimal absorption by physiological absorbers such a themoglobin or water. The absorption and emission spectra in aqueous solution were found to be 674 mn/692 mm and the c = 2.2 x 105 M-Lorn. I. The molecular weight as calculated by L/CMass was 143.0.4 for GCFH28/8010755; found 1431.5 Media.

Property	Specification
Absorbance and emission spectrain 1x PBS	[]
MW	1432 g/mol
Fluorescence excitation	675 nm
Fluorescence emission	693 nm
Absorbance	675 nm± 5 nm
Appearance	Dark blue solid, soluble in water or aqueous buffer

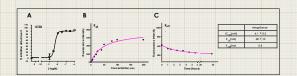
Absorbance and fluorescence maxima of IntegriSense 680 in 1X PBS

3 In Vitro Binding

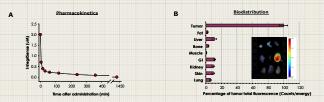
A: IC₀, IHEX732 cells stably transfected with a, II, IEEE/25-a, III, were incubated in the presence of varying concentration of Integrisense at 49 of for 30 min. Cells were tryptimized, washed and added to trionochi-cnoated microtite relels, and allowed to attach at 37°C for 2 hr; non-attached cells were gently washed away. Attached cells were quantified by colorimetric detection of hexaminidase enzymatic activity in a micropalte reader.

B. K_c: HEX293-α,β₃ cells were incubated with varying concentrations of IntegriSense as described above. The amount of agent bound to integrins on HEX293-α,β₃ cells was determined by flow cytometry. Data was analyzed using Flowlo software and K_d values calculated using SigmaPlot 10.

C. K., EEC23-a, B, were incubated with 100 nM Integrisense at 49°C for 30 min, transferred into PBS containing 10 mM of unlabeled compound (parent compound). The amount of agent bound to integrins on HEC23-a, B, cells was determined by flow cytometry before mixing with parent compound and at various times after mixing. Data was analyzed using FlowJo software and Ky, values calculated using SigmaPot 10.



4 Pharmacokinetic and Biodistribution Profile

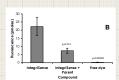


A. BALB/c mice were injected i.v. with IntegriSense 680 (2 nmoles). Blood was collected at various times, plasma obtained by centrifugation, and fluorescence read using a fluorescence increplate reader. B. Human riabdomyosarcoma A673 tumor-bearing nucle mice were injected i.v. with IntegriSense (2 nmoles) and scarfided 24 hrs later. Organs were excised and imaged on the FMT 2500th quantitative tomography system using the reflectance mode. Regions of interest (ROI) were drawn around each organ using the FMT software and the mean fluorescence (Counts/Energy) determined for each organ and normalized to the mean fluorescence (to the count of the tumor (set to 100%). Shown are Mean ± SEM. Insert shows a representative image of the fluorescence (detected in different organs. * tumor.

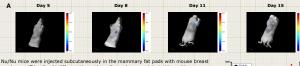
5 Integrin-Targeted Agent Specifically Detects Tumor-Associated Integrins: Quantification with FMT In Vivo Imaging



Nu/Nu mice were injected subcutaneously bilaterally in the mammary fat pads with human rhabdomyosocroma AG73 cells. Mice were nandomized according to tumor volume and injected i.v. with 2 moiles of Integrisense in the absence or presence of the unlabeled parent compound (100 moiles) which acts as a competitor, and imaged 24 hrs later by PRT. Control mice were injected with 2 moiles of free dye. In Appearance of the properties of the propert



6 IntegriSense Signal Strongly Correlates with Tumor Volume

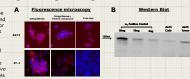


Nu/Nu mice were injected subcutaneously in the mammary fat pads with mouse breast carcinoma 4T1 cells. At different times thereafter, mice were injected iv. with IntegriSensee and imaged 24 hrs later by FMT. A. Representative volume rendering integrishes and imaged 24 hrs later by FMT. A. Representative volume rendering a subcomparing the subcomparing times and integrishes are subcompared to the subcomparing times. The subcomparing time in the subcomparing time calculated as mm² = (length x width²)/2. Images were reconstructed using the FMT software and the total amount of fluorescence (prom) was quantified in specific ROIs around each tumor. A strong correlation was seen between tumor volume and IntegriSense signal (x² = 0.87).

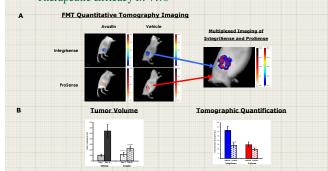
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7 Specific Binding of Integrin-Targeted Agent to Tumor Cells

A Tunor calls (A673 or 47-1) were cultured in the presence of IntegriSense (20 mM), Integrisense and presence of IntegriSense (20 mM), Integrisense and 10 min at 37%. Calcare common control of the common control of the common control of the contr



8 Integrin-Targeted Imaging Agent can be used to Assess Therapeutic Efficacy In Vivo



AG73 Lumor-bearing mice (implanted in the flank) were randomized into 2 groups: Avastin or Vehicle. Mice in the vastin group received 2 mg/kg Avastin (bevacrumab, Generlech, CA) i.p. 2x per week, while mice in the Vehicle group received Avastin Seven days later, mice were injected i.v. with 2 moles IntegriSense and 2 mnoles Prosense*750 (PerkinEimer), and were imager 24 hrs later.

A. Representative isosurface rendering projections of a mouse treated with Avastin and a mouse treated with vehicle. Note the differential localization of IntegriSense (blue) and ProSense (red) within the same tumor. B. Avastin significantly inhibited tumor growth as assessed by calculated tumor volume derived from caliper measurements (left). A significant decrease in IntegriSense signal of 63% (p=0.07), but not ProSense signal (p=0.13) was observed 1 week after treatment (right).

9 Summary

including cancer, and as such represent viable biomarkers for the progression of these diseases. We have developed IntegriSerise 680s, an integrin-targeted molecular imaging agent that allows the non-imassive imaging of disease status and progression. In breast and rhabdomyosacroma tumons, this agent detects the integrin av30 localized in the tumor. Pairing of an integrin antagonist treatment with IntegriSense 680 provides a mechanistic biomarker approach for assessing target coverage. Furthermore, treatment with Avastin showed quantitative changes in integrin imaging with as little as 1 week of treatment. The ability to spatially and temporally visualize and quantify tissue integrin levels in vivo using this targeted fluorescent agent and quantitative FMT tomographic imaging will greatly improve the ability to assess integrin expression during tumor development and metastasis, to develop novel anti-integrin therapies, and to monitor treatment efficacy longitudinally.

10 References

Coleman PJ et al. Nonpeptide α, β_i Antagonists. Part11: Discovery and preclinical evaluation of potent α, β_i antagonists for the prevention and treatment of osteoporosis. J. Med. Chem. 47, 4829-4837 (2004).

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